## AMENDMENTS TO THE SPECIFICATION:

**Please amend the paragraph beginning at page 5, line 22, as follows:**

5'primer: GGAATTCGAACGCTGACGTCATCAACCCG (SEQ ID NO: 1)

**Please amend the paragraph beginning at page 5, line 23, as follows:**

3' primer: GAAGATCTGTCTCATACAGAACTTATAAGATTCCC (SEQ ID NO: 2)

(matation one: three (3) nucleotides just upstream of Bgl II site was deleted in order for transcription to start from proper position after the insertion of the AAAAA sequence according to described beneath) Clone the PCR product in between EcoR I-BGL II, into the original pBLUESCRIPT II KS-H 1 (Brummelkamp T R et al. cited above) vector, verify the plasmid DNA by sequencing:

**Please amend the paragraph beginning at page 5, line 29, as follows:**

The modified sequence: (SEQ ID NO: 3)

**Please amend the paragraph beginning at page 6, line 15, as follows:**

2. Construction of the vector with mutated DUAL-H 1 promoters (here below referred to as pDH, stands for plasmid with Dual H 1 promoters) PCR amplify the fragment between EcoR I-BGL II of the above modified vector, with the following primers:

5'primer: ACGCGTCGACGAATTCGAACGCTGACGTCATCAACCCG (SEQ ID NO: 4)

3'primer: CCCAAGCTTGTCTCATACAGAACTTATAAGATTCCC (SEQ ID NO: 5)

**Please amend the paragraph beginning at page 6, line 27 to page 7, line 4, as follows:**

3. Put the Renilla luciferase target sequence into pDH to form PDHRL: A sequence corresponding to nt 82-100 of Renilla luciferase MRNA was used as the test DNA. siRNA targeting this site of the Renilla luciferase was known to be active (Brummelkamp T R et al. cited above). Two oligo DNA were synthesized and annealed to each other to make the double-stranded DNA:

5'GGGGAAGATCTAAAAAAATAAATGAATCAAGAACATTTTTAAGCTTGG GG (SEQ ID NO: 6);

5'CCCCAAGCTTAAAAATGTTCTTGATTCATTTATTTTTTTAGATCTTCCCC (SEQ ID NO: 7) The above double stranded DNA was cleaved with Bgl II-Hind III and cloned in between Bgl II-Hind III sites in pDH. Verification of the correct insertion of the DNA fragment into the plasmid DNA was done by cleavage by Bgl II+SAL I digestion, where the correct clone should give rise to A-250BP fragment. All three clones tested showed to have the correct insert (FIG. 2)

**Please amend the paragraph beginning at page 7, line 27, as follows:**

19-mer randomized region GGGGAAGATCTAAAAA

NNNNNNNNNNNNNNNNNNN TTTTTAAGCTTGGGG (SEQ ID NO: 8)

20-mer randomized region GGGGAAGATCTAAAAA

NNNNNNNNNNNNNNNNNNNNTTTTTAAGCTTGGGG (SEQ ID NO: 9)

21-MER randomized region GGGGAAGATCTAAAAA

NNNNNNNNNNNNNNNNNNNNN TTTTTAAGCTTGGGG (SEQ ID NO: 10)

**Please amend the paragraph beginning at page 7, line 32, as follows:**

The oligonucleotides were allowed to anneal to a primer CCCCAAGCTTAAAAA (SEQ ID NO: 11) and filled in with Klenow fragment in the presence of 1 mM concentration of dNTP in proper buffer (all chemicals other than DNA oligonucleotides were purchased from New England Biolabs Inc. unless otherwise specified). The duplex oligos were cleaved with Bgl II-Hind III and cloned in the Bgl II-Hind III sites of the pDH to form PDH-LIBRARYA.

**Please amend the paragraph beginning at page 8, line 8, as follows:**

AAAGGGTTTACGTGGTTGG (SEQ ID NO: 12) AATCGTCTTATTTGCATGC (SEQ ID NO: 13)

AATTGACATGTGAGCTTGG (SEQ ID NO: 14) AGTAGCTTGTTGAGGTTGG (SEQ ID NO: 15)

CAGCATCACTGTATGTGTC (SEQ ID NO: 16) CTATCTTCGTGGAGGTTGG (SEQ ID NO: 17)

CTATGAAGGTGGTGATGCG (SEQ ID NO: 18) CTTAATTGGTGGTTGTAGG (SEQ ID NO: 19)

TGGCTGTATGTGAGTGGCT (SEQ ID NO: 20) TTAATCTCTGGTGTCCTAA (SEQ ID NO: 21)

TTGTAGGGACTTGGATGAT (SEQ ID NO: 22)